

THE CLSU INTERNATIONAL JOURNAL OF SCIENCE & TECHNOLOGY www.clsu-ijst.org



Species Listing of Macrofungi Found in Paracelis Mountain Province, Philippines

Angeles M. De Leon^{1,2*}, Mariane Ann Y. Pagaduan¹, Bryan E. Panto¹ and Sofronio P. Kalaw^{1, 2}

¹Department of Biological Sciences, College of Science, Central Luzon State University, Science City of Muñoz, Nueva Ecija, Philippines

²Center of Tropical and Mushroom Development Center, College of Science, Central Luzon State University, Science City of Muñoz, Nueva Ecija, Philippines

Email for correspondence: angelesdeleon@clsu.edu.ph

Submitted March 24, 2021. Accepted July 21, 2021. Published online August 31, 2021

Abstract

Mushrooms are an important natural source of food and medicine. In the Philippines, only a few studies have been conducted on the diversity of mushrooms especially in the mountainous areas. The present study was conducted to document the species of macrofungi found in Paracelis, Mountain Province. The knowledge gained from this study can reveal their importance to the community. A total of 37 macrofungi belonging to 16 families, 26 genera, and 29 species were collected and identified. Twenty nine of the collected macrofungi were identified up to its species level and eight were only identified at its genus level. The collected samples were subjected to morphological identification based on its macroscopic and microscopic characteristics. The identified macrofungi were: Auricularia auricula-judae, Conocybe arrhenii, Coprinellus disseminatus, Coprinus sp., Crepidotus mollis, Daldinia concentrica, Earliella scabrosa, Favolus acervatus, Fomes fomentarium, Ganoderma applanatum, Ganoderma fornicatum, Ganoderma lucidum, Hygrocybe sp., Irpex lacteus, Lentinus strigosus, Lenzites elegans, Lepiota lilacea, Lepiota sp., Marasmiellus ramealis, Microporus xanthopus, Mycena sp., Panellus mitis, Paneolus cyanescens, Parasola plicalitis, Psathyrella candolleana, Psathyrella sp., Russula sp. Schizophyllum commune, Trametes elegans, Trametes gibbosa, Trametes hirsuta, Trametes versicolor, Trametes sp. 1, Trametes sp. 2, and *Xylaria papulis.* Out of these macrofungi, four species were identified as edible, viz: Auricularia auricula, Lentinus strigosus, Coprinus disseminatu, and Schizophyllum commune.

Key Words: diversity, edibility, morphology, mushroom, substrate

Introduction

Mushrooms are macrofungus with a distinctive fruiting body which can be either epigeous or hypogeous and large enough to be seen by the naked eye and to be picked by hand (Chang & Miles, 1992; Chang, 1996; Hawksworth, 2001). They grow best during the rainy season and can be found in almost all types of substrates, e.g., soil, grassy ground, rotten wood, decaying organic matter, etc. (López-Quintero et al., 2012). Mushrooms are generally regarded as healthy foods, poor in calories and in fat, but rich in fiber, proteins, chitin, vitamins, minerals, and bioactive mycochemicals (Kalač 2013). Thus, the nutritional and medicinal properties of macrofungi are well-known and documented. In fact, mushrooms have shown several biological activities such as anti-hypertensive (Eguchi et al., 2014), antimicrobial (Reyes et al., 2017; Gaylan et al., 2018), anti-thrombotic (San Pedro et al., 2005).

Furthermore, the number of macrofungi in the Philippines is abundant and its estimated species are between 10,000 up to 25,000 (Mueller et al., 2007). The study of Tadiosa et al. (2007, 2011) on Mt. Cuenca in Batangas and Bazal-Baubo in Aurora reported a total estimated list of 3,695 species and 818 genera of different fungal diversity. Recently, Liwanag et al. (2017) reported that in Norzagaray, Bulacan, a total of 21 species belonging to 10 families and 16 genera were identified. Moreover, Dulay & Maglasang (2017) reported that there were a total of 30 species belonging to 18 families and 21 genera from Bambanaba, Cuyapo, Nueva Ecija. Furthermore, Sibounnavong et al. (2008) documented 7 identified species with 1 species belonging to Order Tulasnellas or Tremellales (Jelly fungi), 4 species belonging to order Polyporales, and 2 species belong to order Agaricales at Puncan, Carranglan, Nueva Ecija. These findings indicate the species richness of mushrooms in the country. However, most researches are concentrated in the agroindustrial areas and very few studies were conducted to document the diversity of macrofungi in the mountainous areas such as the Mountain Province.

Mountain Province is known to be the Philippines biggest and highest chain of mountains (Cordillera Administrative Region [CAR], 2017). It lies within the center of the Grand Cordillera, and is bound to the north by Kalinga, south by Benguet and Ifugao, to the east by llocos Sur, and to northwest by Abra. The province exhibits Type I climate which has two marked distinct seasons: dry and wet. Dry season is from November to April and wet season is from May to October (CAR, 2017). Due to this climatic conditions, macrofungi are abundant within the area, which makes it a very good site for mushroom documentation. Thus, this study was conducted with the aim to collect, identify, document, and characterize the different species of macrofungi that are found in Paracelis, Mountain Province. This study, will help reveal the importance of the macrofungi in the area, and contribute in the growing knowledge in the field of fungal diversity in the country.

Materials and Methods

Ethics Requirement

A letter of request was given to the Mayor of Paracelis, Mountain Province as well as to the barangay chieftain of Poblacion to allow the researchers to conduct the research in their area.

Collection Site

Macrofungi were collected from the Barangay Poblacion of Paracelis, Mountain Province, Philippines in January of 2019. The study sites were thoroughly searched to locate different macrofungi species.

Collection of Samples

The collection was done with the assistance of some of the members of the community who are more familiar with the area. Purposive sampling method was used in the collection of macrofungal species. Macrofungi were collected on their fruiting stage and was photographed in their natural habitat. Other important information were also taken.

The fruit bodies in soil and ground litter were dug carefully so as not to damage their bases. For wood-rotting mushrooms, these were cut off from the bark of trees where they were attached. All collected specimens were then labeled, wrapped in wax paper, placed in paper bags, and brought to the laboratory for identification.

Morphological Characterization

All collected macrofungi were identified based on their macroscopic (fruiting bodies) and microscopic (spore morphologies) features. Morphometric data collected for each of the specimens were the different features of the cap, gills, and stalk of the mushrooms. A spore print was also prepared from fleshy mushrooms, while tissues sectioning was done for non-fleshy mushrooms. This was to observe for microscopic features such as spore color, shape, and size.

To examine the spore under the microscope, the hymenium was cut into pieces and submerged in distilled water, yielding a spore suspension. A few drops of the spore suspension were place on a microscopic slide and covered with a glass cover slip and the mounted spore suspension was examined under the microscope and each spore of the specimen was measured using a micrometer.

Identification of the collected samples was compared to the morphologies of the published literatures of Rydin et al. (1997), Quimio (2001), and Lodge et al. (2004). Taxonomic classification was based on the work of Laessoe and Lincoff (1998), Kuo (2015), Phillips (2006), and Quimio (2001).

The substrate pH was noted by collecting a portion of the substrate (leaf litter, soil, wood bark). The soil and leaf litter substrate were submerged in distilled water and the pH was measured using a pH meter. For the wood substrate, it was soaked overnight in distilled water before measuring the pH of the substrate.

Species Listing

The genera and species of macrofungi that were observed in the area were listed and the number of each species and genera was noted. The natural habitat of the macrofungi species was documented, especially, their locations and their substrates.

Results and Discussion

Study Site

Paracelis is located at the easternmost of Mountain Province with a latitude of 17°10′21.7″ N and longitude of 121°21′58.3 E (Figure 1), with an elevation of 238 meters above sea level (masl), air temperature at 26°C, and 60% relative humidity.

The province has two general types of climate. The western portion is under Type I with two pronounced seasons; dry from November to April and wet during the rest of the year. The eastern portion is under Type II with season not very pronounced: relatively dry from November to April and wet for the rest of the year (Department of Interior and Local Government in Cordillera Administrative Region [DILG CAR], 2015a). The topography of Mountain Province is roughly characterized by very steep slopes and deep ravines. Small patches of gentle stones may be found along river banks, narrow valleys, and tops of ridges. Elevation ranges from 500 meters to as high as 2,714 meters (DILG CAR, 2015b).

The total land area of the municipality of Paracelis is 55,160 has as per record of the DENR-CAR. Only 4,686 hectares or 8.5% of the total land area is classified as alienable and disposable. The bigger part which is, 50,474 hectares is designated as public forest. Woodland comprises the biggest area (25,851 hectares) followed by grassland/shrubland (23,041 hectares). Agricultural areas consist of 5,748 hectares or 10.42% of the total land area (DILG CAR, 2015a). Built-up areas consist of 350 hectares or 0.63% and the riverwash is 170 hectares or 0.31% of the total land area. Forest abounds in the northern and western part but is being denuded due to unabated logging and swidden farming. The total built-up and agricultural areas of 6,098 hectares exceed the 4,686 hectares designated as alienable and disposable. This is due to the fact that the main source of livelihood of the people is agriculture (DILG CAR, 2015a).

Natural Habitat of Wild Mushrooms

Most of the macrofungi grow in decaying woods and logs which are commonly seen in the mountainous place. Macrofungi can suitably occur in most terrestrial habitats, from woodlands to grasslands, however they are most likely found diverse in forests, where they can have the right climate to form mature fruiting bodies. Particularly, moisture to allow their spores to develop (Orgiazzi et al., 2016). Liwanag et al. (2017) documented 11 species of fungi in wood areas where large trees, shrubs, and lianas abound. Furthermore, De Leon et al. (2019) mention that the Philippines supports a very rich and diverse assemblage of macrofungi that are naturally growing on different substrates such as leaf litter, decaying

plant residues, and decomposing logs of trees during the rainy season. As one-third of the fungal diversity is believed to exist in the tropical region, macrofungi putatively comprise 10% of the total fungal diversity (Mueller et al., 2007; Rossman, 1994).

Figure I

Satellite image of Paracelis, Mountain Province.



Note: Paracelis, Mountain Province (n.d.). https://www.google.com/maps/place/Paracelis,+Mountain+Province

Species Profile and Listing

Identified macrofungi were arranged and listed alphabetically according to their Family. Descriptions such as taxonomic name, macrofungi color, size, substrate, and substrates pH were also included. Selected macrofungi is presented in Figure 2.

Agaricaceae

Lepiota lilacea

<u>Description</u> :pileus is bell-shaped and size is 20–30 mm in diameter with patches over the surface. The scaly stipe length is 25–35 mm long and 3.5 mm thick. The gills are free, close, and white color. Spores 4.65 x 2.86, ellipsoid and spore print is white. <u>Edibility : inedible</u> <u>Substrate : decaying wood</u>

<u>pH: 6.80</u>

Auriculariaceae

Auricularia auricula

<u>Description</u> :commonly known as "Tengang Daga". It is lobed and veined like ear shaped, has a viscid surface, smooth, wavy margin, and brown-colored. It measures 43 mm wide and 18 mm long. Spores $14 \times 5.5 \mu m$ and sausage-shaped. <u>Edibility</u> : edible

Substrate : decaying wood

<u>рН: 6.17</u>

Bolbitaceae

Paneolus cyanescens

<u>Description</u> :has a convex pileus, light brown to light gray color, surface is smooth and measures 18.25 mm diameter. Gills attachment is adnexed, close and gray in color. Stipe measures 118 mm long by 2.8 mm thick. Spores elliptical and smooth, black. <u>Edibility</u> : psychoactive, medicinal

Substrate : dung

<u>pH: 6.17</u>

Conocybe arrhenii

<u>Description</u> :brick red at center, convex pileus measures 23 mm in diameter with striate margin. Gills are crowded and adnexed. Stipe is pale brown at the base, becoming paler towards top and has a length of 42 mm long and 2.3 mm thick. Stipe surface is scaly with white ring on top. Spores ellipsoidal smooth 7.5 x 4 μ m, brown. Edibility : inedible

Substrate : decaying banana

<u>pH: 7.35</u>

Coprinaceae

Coprinus sp.

<u>Description</u> :pileus is campanulate with a measurement of 10 mm in diameter. Gills are free to adnexed and deliquescent in age. Stipe is thin and measures 4 mm long and 1.5 mm thick. Spores 7 x 9.5 μ m, narrowly elliptical, spore print black.

Edibility : inedible

Substrate : dung

<u>pH: 6.45</u>

Crepidoptaceae

Crepidotus mollis

<u>Description</u> :convex pileus covered with dark brown, small fibrous scales, measures 15–20 mm broad and 20–23 mm wide. Gills are close, narrow, brown to red-brown. Stipe is absent, merely a short basal plug covered with minute hairs. Spores broadly ellipsoidal, smooth, 7–9 x 5–7 μ m, brown in color. Edibility : inedible

Substrate : decaying log

<u>pH: 7.46</u>

Fomitopsidaceae

Fomitopsis feei

<u>Description</u> :semi-circular to fan-like shaped fruiting body with a measurement of 82.9 x 43.5 mm. The upper surface is purple in color with a white margin. Spores 5– 6.5 x 2–3.5 μ m, cylindrical to oblong in shape. <u>Edibility</u> : medicinal

Substrate : decaying log

<u>pH: 7.41</u>

Ganodernataceae

Ganoderma applanatum

<u>Description</u> :has a hard and tough fruiting body, inverted fan-like structure, brown to dark-brown in color with a white margin. Pileus measures 10–30 cm across, 8–14 cm deep, semicircular in outline, or irregular. Spores 6–9 x 4–5 μ m, ellipsoid, brown in color.

Edibility : medicinal Substrate : decaying log pH: 7.19

Ganoderma fornicatum

<u>Description</u> :fan-like to semicircular shape fruiting body coloring from red brown to brown toward the margin, measures from 2.5 x 3.5 mm up to 28 mm thick at the base.

<u>Edibility : inedible</u> <u>Substrate : decaying wood</u> <u>pH: 7.19</u>

Ganoderma lucidum

<u>Description</u> :with a shiny surface with a measurement of 112.4 in length and 21.8 mm wide. Fruiting body is brown-red to black in color. Spores 8.2–12.1 x 4.8–8.9 μ m, brown.

Edibility : medicinal Substrate : log pH: 7.19

Hygrophoraceae

Hygrocybe sp.

<u>Description</u> : has a red cap with yellow margin and measures 12.5 mm wide and 2 mm long, smooth, and fibrillose. Pileus margin is crenate and slightly incurved. Gills are free and close in spacing. Stipe is clavate 1.5 mm wide and 20 mm tall. Stipe surface is smooth. Spores are large smooth and inamyloid with white spore print.

<u>Edibility : inedible</u> Substrate : soil

pH: 7.40

Irpicaceae

Irpex lacteus

<u>Description</u> :irregular with tooth-like patch white pileus measuring 53.6 x 22.3 mm. It is white when fresh and whitish-yellow to whitish-green when dry. Spores 5–7 x 2–3 μ m, smooth, elliptical, and white spore print.

Edibility : inedible

Substrate : dead log pH: 7.39

Lepiotaceae

Lepiota sp.

<u>Description</u> :pileus is broad, brown with scaly surface and the center remaining darker, margin is finely wavy and straight. Gills attachment is free and equal. Stipe measures 35 mm tall and 5 mm wide and is equal in shape, has a brown smooth

De Leon et al., 2021

surface and a ring on the upper portion. Spores 5–8 x 2.5–4 μ m, subfusiform, smooth, white spore print. <u>Edibility : inedible</u> <u>Substrate : soil</u> <u>pH: 5.40</u>

Polyporaceae

Earliella scabrosa

<u>Description</u> :fan-shaped bracket macrofungi, has a rigid cap with a concentric zone of brown, red, yellow and white measuring from 35.9 x 31 mm. Spores 8–12.5 × 3.5–5 μ m, cylindrical to oblong ellipsoid.

Edibility : inedible

Substrate : decaying logs

<u>pH: 7.19</u>

Favolus acervatus

<u>Description</u> :fan-like shape, light brown fruiting body, with a measurement of 53.6 x 22.3 mm. It has leathery to leathery cork in dried condition. Spores 7–10.5 x 2.5–4 μ m, cylindrical, smooth.

Edibility : inedible Substrate : decaying logs pH: 7.39

Fomes fomentarium

<u>Description</u> :semi-circular, tough and smooth surface, bracket fungi with a measurement of 52.1 x 48.1 mm. Brown-green to brown fruiting body. Spores 12–20 x 4–7 μ m, cylindric, inamyloid, smooth.

<u>Edibility : medicinal</u> <u>Substrate : tree trunk</u> pH: 7.37

Lentinus strigosus

<u>Description</u> :has an uplifted pileus, measures 13–17 mm long and 15–18 mm wide, tomentose margin, hairy surface, clavate shape and hollow flesh. Gills are crowded, decurrent, light brown in color and with smooth margin. Stipe is off-center. Spores measures 11.1 x 7.52 μ m, elliptical in shape, and white color.

Edibility : edible, medicinal

Substrate : decaying wood pH: 6.80

Lenzites elegans

<u>Description</u> :circular to semicircular smooth fruiting body, brown wart pileus with lobed thin margin that measure 81 x 33.6 mm. White to pale cream in color. Spores $5-7 \times 2-3 \mu m$, smooth, cylindric to long elliptic, white spore print.

Edibility : inedible

Substrate : decaying log pH: 7.58

Microporus xanthropus

Description : funnel-shaped fruiting body with light-brown to dark-brown strips that measures 20.8 x 16.7 mm. Smooth and shiny when fresh and dull when dry. Edibility : inedible Substrate : decaying tree branch pH: 6.38

Pycnoporus sanguineus

Description :fan-shaped pileus with a wavy margin that measures 113.7 x 74 mm. Fruiting body is dry, red orange to orange in color. Spores of $5-6 \times 2-3 \mu m$, smooth, hyaline, non-amyloid, thin-walled.

Edibility : medicinal Substrate : dead trunk pH: 6.76

Trametes elegans

<u>Description</u>: irregular to semi-circular, thick, rough/lumpy to smooth pileus towards the margin that measure 29.9 x 35 mm. White and cream when fresh and pale when dried. Spores 5–7 x 2–3 μ m, smooth, cylindric to long elliptic, white spore print. Edibility : inedible

Substrate : decaying trunk

pH: 7.21

Trametes gibbosa

Description: has a semi-circular pileus that measures from 61.4 to 40 mm. It has wavy margin with dirty white color. Spores ellipsoidal to sausage-shaped, smooth, 4-6 x 2-2.8µm; hyaline, inamyloid, white spore print.

Edibility : medicinal Substrate : living tree pH: 7.20

Trametes hirsuta

Description :circular to semi-circular fruiting body with a hairy surface with a concentric color of yellow-green to green to brown that measures 81 x 33.6 mm. Spores cylindrical, smooth, $5.5-8.5 \times 1.6-2.5 \mu m$; inamyloid, white spore print. Edibility : medicinal Substrate : dead tree trunk

pH: 7.58

Trametes versicolor

Description :fan-like shaped pileus which measures 113.7 x 74 mm, margins velvety surface with folds and light brown to dark-brown color arranged in alternating pattern. Spores curved cylindrical (sausage-like), smooth, $4-6 \ge 1.5-2.5 \mu m$; inamyloid, white spore print.

Edibility : medicinal Substrate : decaying logs pH: 8.03

Trametes sp. 1 and *Trametes* sp. 2

<u>Description</u> :fan-shaped to semi-circular pileus with smooth margin and measures 22.2 x 27.2 mm. The fruiting body is dry, black to brown (*Trametes* sp. 1) and green to dirty white (*Trametes* sp. 2). Spores 4.5–5.5 x 1.5–2 μ m, cylindric, smooth, inamyloid. <u>Edibility : inedible</u> Substrate : dead trunk

<u>pH: 7.75</u>

Psathyrellaceae

Coprinellus disseminatus

<u>Description</u> :pileus measures 9–11 mm across, radially fibrillose surface and plicatestriate margin. Gills that are closely crowded. Stipe is 5–25 mm long and 1–2 mm thick, smooth surface towards the base, equal shape and hollow flesh that is centrally attached to the pileus. Spores 8.53 x 4.50 μ m, ellipsoid with a pore and black color. Edibility : edible

Substrate : decaying woods

<u>pH: 6.18</u>

Parasola plicalitis

<u>Description</u> :has a conical pileus, smooth surface and pleated, measurement is 10–15 mm in diameter. Gills are free and close or nearly distant. Stipe is smooth and equal, fragile, white color without a ring and measures 35–40 mm and 1 mm thick. Spores irregularly ellipsoidal to ellipsoidal, 10–13 x 7–10.5 x 6.5–8 µm, black spore print. <u>Edibility : inedible</u>

Substrate : decaying banana trunk pH: 7.40

Psathyrella candolleana

<u>Description</u> :convex pileus, center has a yellow-brown color and the rest is cream white which measures 17 mm in diameter, smooth cap surface and margin. The gills are adnexed, brown color, and crowded in spacing. Stipe is equal and has a length of 35 mm and 2.1 mm thick, smooth in surface. Spores, ellipsoidal, smooth, 6.5–9.5 x 4– 5 μ m, spore print dark brown.

Edibility : inedible

<u>Substrate : decaying banana leaf</u> pH: 7.50

Psathyrella sp.

<u>Description</u>: pale brown becoming cream in the center of the pileus, measures 15–20 mm wide, texture is smooth but slightly viscid. White gills are cream to tan and slightly decurrent. Stipe is smooth to cream white which measures 20–40 mm tall and 1–2 mm wide. Spores 6.5–9.5 x 4-–5 μ m, ellipsoid, smooth, dark brown in color.

Edibility : inedible

Substrate : decaying wood

<u>pH: 6.20</u>

Russulaceae

Russula sp.

<u>Description</u> :has a depressed violet pileus with smooth surface and measures 9.8 mm wide and 8.9 mm long, margin is smooth with crowded gill spacing. Stipe measures 5.82 mm long and 3.51 mm wide and it is equal and no ring and volva. Spores 6.5–8.5 x 5.5-8 μ m, usually globose or subglobose, cream to pale yellow spore print. <u>Edibility : inedible</u> <u>Substrate : decaying wood</u>

pH: 6.70

Schizophyllaceae

Schizophyllum commune

<u>Description</u>: locally known as "kudit." Pileus measures 23 mm across, with a whitish gray color when dry or brown color when moist, surface is felty and scurfy with crenate and straight margins with streaked surface. Gills is crowded. Spores cylindrical to ellipsoidal, smooth, 4–6 x 1.5–2.5 μ m, white spore print.

Edibility : edible

Substrate : decaying wood pH: 6.65

Tricholomataceae

Mycena sp.

<u>Description</u> :has a small conical pileus, dry with a measurement of 4 mm wide and 3 mm tall. Gills attachment are adnate and close gills spacing. The thin and equal stipe is 5 mm tall and 1 mm wide. Spores cylindric to long-ellipsoid, 2.5–3.5 μ m. <u>Edibility</u> : edible <u>Substrate</u> : banana leaf <u>pH</u>: 7.01

Panellus mitis

<u>Description</u> :has a shell-shaped pileus that measures 10-20 mm in diameter with pale brown color, surface is smooth and has a radiating margin. The white gills are well-spaced. Spores 3.5-6 x 1–1.5 μ m, cylindrical to sausage shape. <u>Edibility</u> : inedible

Substrate : dead trunks pH: 7.46

Marasmellus ramealis

<u>Description</u> :cream to pale brown convex-shaped pileus with a diameter of 10–15 mm, surface texture is wrinkled, and margin is lobe and straight. Gill attachment is adnexed and broad gills are widely spaced. Stipe is tapered downward and clavate with a length of 10_15 mm and 0.5 mm thick with smooth surface. Spores, elongate, elliptical, $8.0-10 \times 3-4 \mu m$.

Edibility : inedible

Substrate : decaying wood pH: 7.20

Xylariaceae

Daldinia concentrica

<u>Description</u> :has a ball shape, hard, shiny purple fruiting body, with a measurement of 32 x 27.6 mm. Reddish-purple to purple when young and becoming black when old and about to dry. Spores ellipsoidal to fusiform, 12–17 x 6–9 μ m, spore print is black. <u>Edibility : medicinal</u>

Substrate : decaying tree pH: 8.00

Xylaria papulis

<u>Description</u> :finger-shaped to club-shaped macrofungi measuring from 85.3 x 50.4 up to 101 x 56.2 mm. It has a yellow fruiting body that turns black when bruised. Spores dark brown, broad fusoid, 7–9 × 4–4.5 μ m. <u>Edibility</u> : inedible <u>Substrate</u> : soil

pH: 7.55

Habitat and Importance of Collected Macrofungi

The collected macrofungi in the study site were classified per taxonomic family and their growth habit as well as their importance were also evaluated. The collected samples belonging to 16 families namely: Agaricaceae, Auriculariaceae, Bolbitaceae, Crepidoptaceae, Coprinaceae, Fomitopsidaceae, Ganodermataceae, Hygrophoraceae, Irpicaceae, Lepiotaceae, Polyporaceae, Psathyrellaceae, Russulaceae, Schizophyllaceae, Tricholomataceae and Xylariaceae. Among the 16 families, Polyporaceae has the highest collected and identified macrofungi with a total of 13 species. Of the 37 macrofungi identified, 21 species are classified as saprobic, 10 medicinal, 2 nutraceutical, and others with 1 species each are therapeutic, bioremediator, and pathogen (Table 1).

Discussion

The abundance of Polyporaceae in this study is understandable since this family is commonly found growing in decaying log, fallen branches of trees, hardwood and tree trunks (Liwanag et al., 2017) that are rich in mountainous regions like the Mountain Province. Similar results were obtained with the study of De Leon et al. (2013) in Tarlac, Pampanga and Zambales, and Tadiosa et al. (2011) in Aurora which are mountainous provinces. This result is not surprising, for polypores play a primary and central role in decomposition and nutrient cycling in forest ecosystems (Adarsh et al. 2018). The collected macrofungi were mostly found growing in wood, decaying tree trunks, and decaying branches of trees. Wooddecaying fungi are important natural components of indigenous forests; their growth and distribution are affected by a multitude factors such as difference in elevations (Adarsh et al., 2018), host specificity (Gilbert & Sousa 2002) and many others. The pattern of host specificity had been previously observed from tropical polypores and are generally most prominent among fungi in decaying wood that are in their early decomposing stages (Gilbert & Sousa, 2002). The colonization of polypores is through germinating spores and migratory mycelia (Adarsh et al., 2018). They are dispersed by airborne basidiospores and geographical barriers, such as mountains, rivers, and floodplains (Gibertoni et al., 2016).

Studies of Clausen-Heilmann and Christensen (2005) and May (1991) suggest that highly diverse forests (with a high number of tree species and scattered trees of each species) would promote low specificity of fungal species. Thus, low diversified forest results to higher fungal species specificity (Gilbert, 2005). Mountain Province is considered as a diverse forest, since it is composed of several species of fruit trees of the agroforestry farms (Baldic, 2016) as well as other naturally occurring tree species on the province. However, Schmitt (2005) mentioned that the macrofungi is not widely distributed as that of the trees; they also added that there must be other factors that limit the distribution of macrofungi. These limits could be the environmental factors that are associated with differences in elevation and more importantly in the temperature, precipitation duration, intensity of illumination, physical and chemical characteristics of the soil, topographic position, and tree density (Zhang et al., 2010).

Second to Polyporaceae is Psathyrellaceae with four species followed by Ganodermataceae, and Tricholomataceae with three species each. De Leon et al. (2013) reported 76 species of macrofungi in six Aeta tribal communities in Luzon. While Corazon and Licyayo (2018) in the Cordillera Region reported that there were 30 species from 23 genera of wild edible fungi identified and most of the species were found in Benguet and Mt. Province areas where the coniferous type of forest was abundant and conducive to the growth of wild edible mushrooms. The exhibited patterns of diversity of macrofungi were relative to their substrates, host availability, and the climatic condition (Parveen et al., 2017).

Twenty one species of the collected marcrofungi were classified as saprobic. These were: *L. lilacea, A. auricularia-judae, C. arrheni, Coprinus* sp., *C. mollis, G. fornicatum, Hygrocybe* sp., *Lepiota* sp., F. acervatus, *L. elegans, Trametes* sp. 1 and 2, *C. plicalitis, Psathyrella* sp., *P. candolleana, Mycena* sp., *P. mitis, M. ramealis, Rusulla* sp. and *X. papulis.* Almost all of the species collected were saprophytic fungi which obtain food by absorbing dissolved organic material from decayed wood and soil organic matters. According to Parveen et al. (2017), saprophytic fungi break down dead and decaying organic matter into simple substance that can be taken up and recycled by plants, thus maintaining forest ecosystem balance. Niem and Baldovino (2015) reported that in Cavinti Underground River and Cave Complex in Cavinti, Laguna, most species were saprotrophs.

Aside from being edible, many of the identified mushrooms were also medicinal such as *P. cyanescens*, G. applanatum, G. lucidum, *F. fomentarium*, *L. strigosus*, *P. sanguineus*, *T. gibbosa*, *T. hirsuta*, *T. versicolor*, *C. disseminatus*, *D. concentrica*. These mushrooms could be significant sources of bioactive compounds with various functional activities. One of the most important medicinal mushrooms is *G. lucidum*. It is used to treat chronic hepatitis, hypertension, arthritis, insomnia, bronchitis, asthma, gastric ulcer, diabetes, and cancer. It possesses anti-tumor activity and has also been found to inhibit platelet aggregation and to lower blood pressure, cholesterol, and blood sugar (Borchers et al., 2004).

De Leon et al., 2021

Selected Species of Macrofungi Collected from Paracelis Mountain Province: A *Lepiota lilacea*. B *Auricularia auricula-judae*. C *Fomitopsis feei*. D *Ganoderma applanatum*. E *Ganoderma fornicatum*. F *Ganoderma lucidum*. G *Irpex lacteus*. H *Earliella scabrosa*. I *Favolus acervatus*. J *Fomes fomentarium*. K *Lentinus strigosus*. L *Lenzites elegans*. M *Microporus xanthopus*. N *Pycnoporus sanguineus*. O *Trametes elegans*. P *Trametes gibbosa*. Q *Trametes hirsute*. R *Trametes versicolor*. S *Trametes* sp. 1. T *Trametes* sp. 2. U *Coprinellus disseminatus*. V *Psathyrella* sp. W *Schizophyllum commune*. X *Daldinia concentrica*.



Table I

The Number of Bracket Macrofungi Listed per Family with their Substrate and Importance

AgaricaceaeL. IllaceaSolitarySaprobic (Kuo, 2015)AuriculariaceaeSaprobic (Kuo, 2015)BolbitaceaeP. cyanescensGregariousSaprobic (Kuo, 2015)C. arrheniiSolitary to gregariousSaprobicC. arrheniiSolitary to gregariousSaprobicCoprinaceaeCoprinaceaeGregarious in clustersSaprobicGanodermataceaeG. applanatumSolitaryMedicinal (De Leon et al., 2013)G. foreiratumSolitaryMedicinal (De Leon et al., 2013)G. formicatumSolitarySaprobicHygrophoraceaeI. lacteusResupinateBioremediation (Novotny et al., 2009)IppicaceaeI. lacteusResupinateBioremediation (Novotny et al., 2009)Lepiota sp.SolitarySaprobicPolyporaceaeE. scabrosaGregariousSaprobicF. fomentariumSolitarySaprobicPolyporaceaeE. scabrosaGregariousSaprobicF. fomentariumSolitarySaprobicJ. acteusSolitarySaprobicPolyporaceaSaprobicJ. acteusSolitarySaprobicPolyporaceaeE. scabrosaGregariousMedicinal (Peintner et al., 1998)L. elegansSolitarySaprobicJ. strigosusGregariousMedicinal (Peintner et al., 2013)M		GROWTH HABIT	IMPORTANCE
L. IliaceaSolitarySaprobic (Kuo, 2015)AuriculariaceaeA. auriculariaGregariousSaprobic (Kuo, 2015)BolbitaceaeP. cyanescensGregariousSaprobicCrepidoptaceaeC. arrheniiSolitary to gregariousSaprobicCoprinaceaeCoprinaceaeCoprinaceaeCoprinaceaeGanodermataceaeG. applanatumSolitaryMedicinal (De Leon et al., 2013)G. lucidumGregarious in clustersSaprobicGanodermataceaeG. applanatumSolitaryMedicinal (De Leon et al., 2013)G. lucidumGregarious in clustersMedicinal (De Leon et al., 2013)G. fornicatumSolitarySaprobicHygrocybe sp.SolitarySaprobicIrpicaceaeLepiota csp.SolitarySaprobicPolyporaceaeLepiota sp.SolitarySaprobicPolyporaceaeLegansSolitarySaprobicF. fomentariumSolitarySaprobicF. fomentariumSolitarySaprobicL strigosusGregariousMedicinal (Peintner et al., 1998)L elegansSolitarySaprobicF. fomentariumSolitarySaprobicF. fomentariumSolitarySaprobicF. fomentariumSolitarySaprobicF. fomentariumSolitary <t< td=""><td>Agaricaceae</td><td></td><td></td></t<>	Agaricaceae		
AuriculariaGregariousSaprobic (Kuo, 2015)Bolbitaceae-P. cyanescensGregariousMedicinal (Bustillos et al., 2014)C. arrheniiSolitary to gregariousSaprobicCrepidoptaceaeC. moliisGregariousSaprobicCoprinaceaeCoprinaceaeFeiGregarious in clustersSaprobicGandermataceaeG. applanatumSolitaryMedicinal (De Leon et al., 2013)G. fornicatumSolitary to gregariousSaprobicG. applanatumSolitary to gregariousSaprobicIppiaceaeHygrocybe sp.Solitary to gregariousSaprobicIppiaceaeHygrocybe sp.Solitary to gregariousSaprobicIppiaceaeI. lacteusResupinateBioremediation (Novotny et al., 2009)LepiotaceaeE. scabrosaGregariousSaprobicPolyporaceaeE. scabrosaGregariousSaprobicF. formentariumSolitary to gregariousSaprobicF. formentariumSolitary to gregariousSaprobicLeipiota ceaeI. lacteusGregariousPathogen (He et al., 2018)F. acervatusSolitary to gregariousSaprobicLeipiota sp.Solitary to gregariousMedicinal (Peintner et al., 1998)L. elegansSolitary to gregariousMedicinal (Peint	L. lilacea	Solitary	Saprobic (Kuo, 2015)
A. auriculariaGregariousSaprobic (Kuo, 2015)BolbitaceaeP. cyanescensGregariousMedicinal (Bustillos et al., 2014)C. arrheniiSolitary to gregariousSaprobicCrepidoptaceaeC. mollisGregariousSaprobicCoprinaceaeContropsidaceaeF. feeiGregarious in clustersSaprobicGanodermataceaeG. applanatumSolitary to gregariousSaprobicG. lucidumGregarious in clustersMedicinal (De Leon et al., 2013)G. lucidumGregarious in clustersMedicinal (De Leon et al., 2013)G. lucidumGregarious in clustersMedicinal (De Leon et al., 2013)G. fornicatumSolitary to gregariousSaprobicHygrocybe sp.Solitary to gregariousSaprobicInfacteusResupinateBioremediation (Novotny et al., 2009)Lepiota sp.SolitarySaprobicPolyporaceaeLepiota sp.Solitary to gregariousSaprobicF. fomentariumSolitarySaprobicF. fomentariumSolitarySaprobicF. fomentariumSolitary to gregariousSaprobicJ. aclegansSolitary to gregariousMedicinal (Pentner et al., 2013)M. xarthopusSolitary to gregariousMedicinal (Cat et al., 2013)M. xarthopusSolitary to gregariousMedicinal (Cat et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Ren et al., 2008) <td>Auriculariaceae</td> <td></td> <td></td>	Auriculariaceae		
BolbitaceaeVertailP. cyanescensGregariousMedicinal (Bustillos et al., 2014)C. arrheniiSolitary to gregariousSaprobicCrepidoptaceaeCoprinaceaeCoprinaceaeCoprinaceaeCoprinaceaeFereiCoprinaceaeGregarious in clustersSaprobicFomitopsidaceaeFereiGregarious in clustersGandermataceaeGregarious in clustersMedicinal (De Leon et al., 2013)G. lucidumGregarious in clustersMedicinal (De Leon et al., 2013)G. lucidumGregarious in clustersMedicinal (De Leon et al., 2013)G. fornicatumSolitary to gregariousSaprobicHygrophoraceaeHygrocybe sp.Solitary to gregariousSaprobicIrpicaceaeII. dacteusResupinateBioremediation (Novotny et al., 2009)LepiotaceaeILepiota sp.Solitary to gregariousSaprobicPolyporaceaeIE. scabrosaGregariousPathogen (He et al., 2018)F. dorentariumSolitary to gregariousSaprobicF. fomentariumSolitary to gregariousSaprobicF. fomentariumSolitary to gregariousMedicinal (Papwal et al., 2013)M. xathopusSolitary to gregariousMedicinal (Chapwal et al., 2013)M. xathopusSolitary to gregariousMedicinal (Chapwal et al., 2013)M. xathopusSolitary to gregariousMedicinal (Chapwal et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Chapwal et al., 2013)P. sanguineusSolita	A. auricularia	Gregarious	Saprobic (Kuo, 2015)
P. cyanescens C. arrheniiGregariousMedicinal (Bustillos et al., 2014)C. arrheniiSolitary to gregariousSaprobicCrepidoptaceae	Bolbitaceae		
C. arrhenilSolitary to gregariousSaprobicCrepidoptaceae	P. cyanescens	Gregarious	Medicinal (Bustillos et al., 2014)
CrepidoptaceaeJene in the second	C. arrhenii	Solitary to gregarious	Saprobic
C. mollisGregariousSaprobicCoprinuesp.GregariousSaprobicFomitopsidaceaeF. feeiGregarious in clustersSaprobicGanodermataceaeGregarious in clustersSaprobicG. applanatumSolitaryMedicinal (De Leon et al., 2013)G. lucidumGregarious in clustersMedicinal (De Leon et al., 2013)G. lucidumGregarious in clustersMedicinal (De Leon et al., 2013)G. fornicatumSolitary to gregariousSaprobicHygrophoraceaeHygrophoraceaeInterceaeI. lacteusResupinateBioremediation (Novotny et al., 2009)LepiotaceaeImplicaceaeImplicaceaeE. scabrosaGregariousSaprobicPolyporaceaeSolitary to gregariousSaprobicF. fomentariumSolitary to gregariousSaprobicF. fomentariumSolitary to gregariousSaprobicF. fomentariumSolitary to gregariousMedicinal (Peintner et al., 1998)L. elegansSolitary to gregariousMedicinal (Tapwal et al., 2013)M. xanthopusSolitary to gregariousMedicinal (Tapwal et al., 2013)M. santhopusSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2006)T. hirsutaSolitary to gregariousMedicinal (Ren et al., 2006)T. hirsutaSolitary to gregariousMedicinal (Ren et al., 2013)T. resicolorGregariousMedicinal (Knezević et al., 2013)Trametes sp. 1Solitary to	Crepidoptaceae		
CoprinaceaeCoprinus sp.GregariousSaprobicFomitopsidaceaeFfeeiGregarious in clustersSaprobicGanodermataceaeGitaryMedicinal (De Leon et al., 2013)G. lucidumGregarious in clustersMedicinal (De Leon et al., 2013)G. lucidumGregarious in clustersMedicinal (De Leon et al., 2013)G. fornicatumSolitary to gregariousSaprobicHygrophoraceaeHygrophoraceaeIn lacteusResupinateBioremediation (Novotny et al., 2009)LepiotaceaeILacteusSolitary to gregariousSaprobicPolyporaceaeIE. scabrosaGregariousPathogen (He et al., 2018)F. acervatusSolitary to gregariousSaprobicF. fomentariumSolitary to gregariousSaprobicI. elegansSolitary to gregariousSaprobicI. strigosusGregariousMedicinal (Peintner et al., 1998)I. elegansSolitary to gregariousMedicinal (Tapwal et al., 2013)M. xanthopusSolitary to gregariousMedicinal (Cta et al., 2008)Solitary to gregariousMedicinal (Cta et al., 2008)T. eigbosaSolitary to gregariousMedicinal (Ren et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Ren et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Ren et al., 2008)T. eigbosaSolitary to gregariousMedicinal (Ren et al., 2013)T. resicolorGregariousMedicinal (Ren et al., 2013)T. resicolorGr	C. mollis	Gregarious	Saprobic
Coprinus sp.GregariousSaprobicFonitopsidaceaeGregarious in clustersSaprobicGanodermataceaeGapdanatumSolitaryMedicinal (De Leon et al., 2013)G. lucidumGregarious in clustersMedicinal (De Leon et al., 2013)G. inciatumSolitary to gregariousSaprobicHygrophoraceaeImage and the second secon	Coprinaceae		
FomitopsidaceaeGregarious in clustersSaprobicGanodermataceaeSolitaryMedicinal (De Leon et al., 2013)G. lucidumGregarious in clustersMedicinal (De Leon et al., 2013)G. fornicatumSolitary to gregariousSaprobicHygrophoraceaeHygrophoraceaeImage: SaprobicHygrophoraceaeImage: SaprobicIn lacteusResupinateBioremediation (Novotny et al., 2009)LepiotaceaeImage: SaprobicIspitaceaeSolitarySaprobicPolyporaceaeImage: SaprobicE. scabrosaGregariousPathogen (He et al., 2018)F. acervatusSolitary to gregariousSaprobicF. fomentariumSolitaryMedicinal (Peintner et al., 1998)L. elegansSolitary to gregariousMedicinal (Peintner et al., 2013)M. xanthopusSolitary to gregariousMedicinal (Tapwal et al., 2013)M. xanthopusSolitary to gregariousMedicinal (Cat et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2006)T. hinsutaSolitary to gregariousMedicinal (Ren et al., 2013)P. sanguineusSolitary to gregariousSaprobicT. reviscolorGregariousMedicinal (Ren et al., 2013)T. elegansSolitary to gregariousSaprobicTrametes sp. 1Solitary to gregariousSaprobicTrametes sp. 2S	<i>Coprinus</i> sp.	Gregarious	Saprobic
F. feeiGregarious in clustersSaprobicGanodermataceae	Fomitopsidaceae		
GanodermataceaeG. applanatumSolitaryMedicinal (De Leon et al., 2013)G. lucidumGregarious in clustersMedicinal (De Leon et al., 2013)G. fornicatumSolitary to gregariousSaprobicHygrophoraceaeHygrocybe sp.SolitarySaprobicIrpicaceaeIII. lacteusResupinateBioremediation (Novotny et al., 2009)LepiotaceaeIILepiota sp.SolitarySaprobicPolyporaceaeIIE. scabrosaGregariousPathogen (He et al., 2018)F. acervatusSolitary to gregariousSaprobicF. fomentariumSolitary to gregariousSaprobicI. elegansSolitary to gregariousSaprobicL. elegansSolitary to gregariousMedicinal (Peintner et al., 1998)L. elegansSolitary to gregariousMedicinal (Tapwal et al., 2013)M. xanthopusSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2006)T. hirsutaSolitary to gregariousMedicinal (Ren et al., 2013)T. rametes sp. 1Solitary to gregariousSaprobicTrametes sp. 2Solitary to gregariousSaprobicPsathyrellaceaeISaprobicC. disseminatusGregariousSaprobicPsathyrellaceaeISaprobicPsathyrellaceaeID. Sol	F. feei	Gregarious in clusters	Saprobic
G. applanatumSolitaryMedicinal (De Leon et al., 2013)G. lucidumGregarious in clustersMedicinal (De Leon et al., 2013)G. fornicatumSolitary to gregariousSaprobicHygrophoraceae	Ganodermataceae		
G. lucidumGregarious in clustersMedicinal (De Leon et al., 2013)G. fornicatumSolitary to gregariousSaprobicHygrophoraceaeHygrocybe sp.SolitarySaprobicIrpicaceaeIrpicaceaeIntervention (Novotny et al., 2009)LepiotaceaeLepiota sp.SolitarySaprobicPolyporaceaeE. scabrosaGregariousPathogen (He et al., 2018)F. acervatusSolitary to gregariousSaprobicF. fomentariumSolitaryMedicinal (Peintner et al., 1998)L. elegansSolitary to gregariousSaprobicF. strigosusGregariousMedicinal (Tapwal et al., 2013)M. xanthopusSolitary to gregariousNoedicinal (Tapwal et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Cat et al., 2008)T. elegansSolitary to gregariousMedicinal (Cat et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2006)T. hirsutaSolitary to gregariousMedicinal (Ren et al., 2013)T. versicolorGregariousMedicinal (Ren et al., 2013)Trametes sp. 1Solitary to gregariousSaprobicTrametes sp. 2Solitary to gregariousSaprobicPsathyrellaceaeUUC. disseminatusGregariousSaprobicPsathyrellaceaeUUC. disseminatusGregarious in clustersSaprobicPsathyrellaceaeSaprobicSaprobicPsathyrellaceaeUUC. disseminatusGregarious in c	G. applanatum	Solitary	Medicinal (De Leon et al., 2013)
G. fornicatumSolitary to gregariousSaprobicHygrophoraceaeIHygrocybe sp.SolitarySaprobicIrpicaceaeII. lacteusResupinateBioremediation (Novotny et al., 2009)LepiotaceaeILepiota sp.SolitarySaprobicPolyporaceaeIIE. scabrosaGregariousPathogen (He et al., 2018)F. acervatusSolitary to gregariousSaprobicF. fomentariumSolitaryMedicinal (Peintner et al., 1998)L. elegansSolitarySaprobicL. strigosusGregariousMedicinal (Tapwal et al., 2013)M. xanthopusSolitary to gregariousMedicinal (Tapwal et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2008)T. versicolorGregariousMedicinal (Knezević et al., 2013)T. versicolorGregariousMedicinal (Tapwal et al., 2013)Trametes sp. 1Solitary to gregariousSaprobicTrametes sp. 2Solitary to gregariousSaprobicPsathyrellaceaeIColitaryC. disseminatusGregariousSaprobicPsathyrellaceaeISaprobicPsathyrellaceaeISaprobicPsathyrella sp.Gregarious in clustersSaprobicPsathyrella sp.Gregarious in clustersSaprobic (Kuo, 2015)	G. lucidum	Gregarious in clusters	Medicinal (De Leon et al., 2013)
HygrophoraceaeJolitarySaprobicIrpicaceaeIrpicaceaeI. lacteusResupinateBioremediation (Novotny et al., 2009)LepiotaceaeImpicaceaeLepiota sp.SolitarySaprobicPolyporaceaeImpicaceaeE. scabrosaGregariousPathogen (He et al., 2018)F. acervatusSolitary to gregariousSaprobicF. fomentariumSolitaryMedicinal (Peintner et al., 1998)L. elegansSolitarySaprobicL. strigosusGregariousMedicinal (Tapwal et al., 2013)M. xanthopusSolitary to gregariousNonedible (Tapwal et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2006)T. hirsutaSolitary to gregariousMedicinal (Ren et al., 2006)T. hirsutaSolitary to gregariousMedicinal (Tapwal et al., 2013)T. versicolorGregariousMedicinal (Tapwal et al., 2016)T. hirsutaSolitary to gregariousSaprobicTrametes sp. 1Solitary to gregariousSaprobicTrametes sp. 2Solitary to gregariousSaprobicPsathyrellaceaeImplementariusSaprobicPulcialitisGregarious in clustersSaprobicP. plicalitisGregarious in clustersSaprobicP. plicalitisGregarious in clustersSaprobic (Kuo, 2015)P. saprobicGregar	G. fornicatum	Solitary to gregarious	Saprobic
Hygrocybe sp.SolitarySaprobicIrpicaceaeI1. lacteusResupinateBioremediation (Novotny et al., 2009)LepiotaceaeILepiota sp.SolitarySaprobicPolyporaceaeIE. scabrosaGregariousPathogen (He et al., 2018)F. acervatusSolitary to gregariousSaprobicF. fomentariumSolitaryMedicinal (Peintner et al., 1998)L. elegansSolitarySaprobicL. strigosusGregariousMedicinal (Tapwal et al., 2013)M. xanthopusSolitary to gregariousMedicinal (Tapwal et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2006)T. hirsutaSolitary to gregariousMedicinal (Ren et al., 2013)T. versicolorGregariousMedicinal (Tapwal et al., 2013)T. versicolorGregariousMedicinal (Ren et al., 2006)T. hirsutaSolitary to gregariousSaprobicTrametes sp. 1Solitary to gregariousSaprobicPsathyrellaceaeIC. disseminatusC. disseminatusGregarious in clustersSaprobicP. plicalitisGregarious in clustersSaprobic (Kuo, 2015)Psathyrella sp.Gregarious in clustersSaprobic (Kuo, 2015)	Hygrophoraceae		
IrpicaceaeI. lacteusResupinateBioremediation (Novotny et al., 2009)LepiotaceaeLepiota sp.SolitaryLepiota sp.SolitarySaprobicPolyporaceaeE. scabrosaGregariousF. acervatusSolitary to gregariousSaprobicF. fomentariumSolitaryMedicinal (Peintner et al., 2018)L. elegansSolitarySaprobicL. strigosusGregariousMedicinal (Peintner et al., 2013)M. xanthopusSolitary to gregariousNonedible (Tapwal et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2006)T. hirsutaSolitary to gregariousMedicinal (Ren et al., 2013)T. versicolorGregariousMedicinal (Tapwal et al., 2018)Trametes sp. 1Solitary to gregariousSaprobicPsathyrellaceaeUUC. disseminatusGregariousNutraceutical (Novakovic et al., 2016)P. plicalitisGregarious in clustersSaprobicP. plicalitisGregarious in clustersSaprobic (Kuo, 2015)P. saprobicGregarious in clustersSaprobic	<i>Hygrocyb</i> e sp.	Solitary	Saprobic
I. lacteusResupinateBioremediation (Novotny et al., 2009)LepiotaceaeElepiota sp.SolitarySaprobicPolyporaceaeF. acervatusSolitary to gregariousSaprobicF. acervatusSolitary to gregariousSaprobicF. fomentariumSolitaryMedicinal (Peintner et al., 2018)L. elegansSolitary to gregariousMedicinal (Peintner et al., 1998)L. elegansSolitary to gregariousMedicinal (Tapwal et al., 2013)M. xanthopusSolitary to gregariousMedicinal (Cota et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2006)T. elegansSolitary to gregariousMedicinal (Tapwal et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Ren et al., 2006)T. elegansSolitary to gregariousMedicinal (Ren et al., 2006)T. elegansSolitary to gregariousMedicinal (Tapwal et al., 2013)T. versicolorGregariousSaprobicTrametes sp. 1Solitary to gregariousSaprobicPsathyrellaceaeC. disseminatusGregarious in clustersSaprobicP. plicalitisGregarious in clustersSaprobic (Kuo, 2015)P. plicalitisGregarious in clustersSaprobic (Kuo, 2015)	Irpicaceae		
LepiotaceaeSolitarySaprobicPolyporaceaeE. scabrosaGregariousPathogen (He et al., 2018)F. acervatusSolitary to gregariousSaprobicF. fomentariumSolitaryMedicinal (Peintner et al., 1998)L. elegansSolitarySaprobicL. strigosusGregariousMedicinal (Tapwal et al., 2013)M. xanthopusSolitary to gregariousNonedible (Tapwal et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2006)T. hirsutaSolitaryMedicinal (Tapwal et al., 2013)T. versicolorGregariousMedicinal (Tapwal et al., 2013)Trametes sp. 1Solitary to gregariousSaprobicPsathyrellaceaeEC. disseminatusGregariousP. plicalitisGregarious in clustersSaprobic (Kuo, 2015)P. sathurella spGregarious in clustersSaprobic (Kuo, 2015)	I. lacteus	Resupinate	Bioremediation (Novotny et al., 2009)
Lepiota sp.SolitarySaprobicPolyporaceaeE. scabrosaGregariousPathogen (He et al., 2018)F. acervatusSolitary to gregariousSaprobicF. fomentariumSolitaryMedicinal (Peintner et al., 1998)L. elegansSolitarySaprobicL. strigosusGregariousMedicinal (Tapwal et al., 2013)M. xanthopusSolitary to gregariousNonedible (Tapwal et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2006)T. hirsutaSolitary to gregariousMedicinal (Ren et al., 2013)T. versicolorGregariousMedicinal (Tapwal et al., 2013)Trametes sp. 1Solitary to gregariousMedicinal (Ren et al., 2006)Trametes sp. 2Solitary to gregariousSaprobicPsathyrellaceaeC. disseminatusGregariousSaprobicP. plicalitisGregarious in clustersSaprobic (Kuo, 2015)P. plicalitisGregarious in clustersSaprobic (Kuo, 2015)	Lepiotaceae		
PolyporaceaeE. scabrosaGregariousPathogen (He et al., 2018)F. acervatusSolitary to gregariousSaprobicF. fomentariumSolitaryMedicinal (Peintner et al., 1998)L. elegansSolitarySaprobicL. strigosusGregariousMedicinal (Tapwal et al., 2013)M. xanthopusSolitary to gregariousNonedible (Tapwal et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2006)T. hirsutaSolitaryMedicinal (Ren et al., 2013)T. versicolorGregariousMedicinal (Tapwal et al., 2013)Trametes sp. 1Solitary to gregariousSaprobicTrametes sp. 2Solitary to gregariousSaprobicPsathyrellaceaeC. disseminatusGregariousSaprobicP. plicalitisGregarious in clustersSaprobicPsathvrella spGregarious in clustersSaprobic (Kuo, 2015)	<i>Lepiota</i> sp.	Solitary	Saprobic
E. scabrosaGregariousPathogen (He et al., 2018)F. acervatusSolitary to gregariousSaprobicF. fomentariumSolitaryMedicinal (Peintner et al., 1998)L. elegansSolitarySaprobicL. strigosusGregariousMedicinal (Tapwal et al., 2013)M. xanthopusSolitary to gregariousNonedible (Tapwal et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2006)T. gibbosaSolitary to gregariousMedicinal (Ren et al., 2013)T. versicolorGregariousMedicinal (Knezević et al., 2018)Trametes sp. 1Solitary to gregariousSaprobicTrametes sp. 2Solitary to gregariousSaprobicPsathyrellaceaeC. disseminatusGregariousSaprobicP. plicalitisGregarious in clustersSaprobic (Kuo, 2015)Psathyrella spGregarious in clustersSaprobic (Kuo, 2015)	Polyporaceae		
F. acervatusSolitary to gregariousSaprobicF. fomentariumSolitaryMedicinal (Peintner et al., 1998)L. elegansSolitarySaprobicL. strigosusGregariousMedicinal (Tapwal et al., 2013)M. xanthopusSolitary to gregariousNonedible (Tapwal et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2006)T. gibbosaSolitary to gregariousMedicinal (Ren et al., 2006)T. hirsutaSolitary to gregariousMedicinal (Knezević et al., 2018)T. versicolorGregariousMedicinal (Tapwal et al., 2013)Trametes sp. 1Solitary to gregariousSaprobicPsathyrellaceaeC. disseminatusGregariousSaprobicP. plicalitisGregarious in clustersSaprobic (Kuo, 2015)P. sathyrella spGregarious in clustersSaprobic (Kuo, 2015)	E. scabrosa	Gregarious	Pathogen (He et al., 2018)
F. fomentariumSolitaryMedicinal (Peintner et al., 1998)L. elegansSolitarySaprobicL. strigosusGregariousMedicinal (Tapwal et al., 2013)M. xanthopusSolitary to gregariousNonedible (Tapwal et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitary to gregariousMedicinal (Ren et al., 2006)T. gibbosaSolitary to gregariousMedicinal (Ren et al., 2006)T. hirsutaSolitaryMedicinal (Knezević et al., 2013)T. versicolorGregariousMedicinal (Tapwal et al., 2013)Trametes sp. 1Solitary to gregariousSaprobicPsathyrellaceaeSolitary to gregariousSaprobicP. plicalitisGregariousSaprobicP. saprobicSaprobicSaprobic	F. acervatus	Solitary to gregarious	Saprobic
L. elegansSolitarySaprobicL. strigosusGregariousMedicinal (Tapwal et al., 2013)M. xanthopusSolitary to gregariousNonedible (Tapwal et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitaryTherapeutic (Awala & Ayetayo,2015)T. gibbosaSolitary to gregariousMedicinal (Ren et al., 2006)T. hirsutaSolitaryMedicinal (Knezević et al., 2018)T. versicolorGregariousMedicinal (Tapwal et al., 2013)Trametes sp. 1Solitary to gregariousSaprobicTrametes sp. 2Solitary to gregariousSaprobicPsathyrellaceaeC. disseminatusGregarious in clustersNutraceutical (Novakovic et al., 2016)P. plicalitisGregarious in clustersSaprobic (Kuo, 2015)	F. fomentarium	Solitary	Medicinal (Peintner et al., 1998)
L. strigosusGregariousMedicinal (Tapwal et al., 2013)M. xanthopusSolitary to gregariousNonedible (Tapwal et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitaryTherapeutic (Awala & Ayetayo,2015)T. gibbosaSolitary to gregariousMedicinal (Ren et al., 2006)T. hirsutaSolitaryMedicinal (Knezević et al., 2018)T. versicolorGregariousMedicinal (Tapwal et al., 2013)Trametes sp. 1Solitary to gregariousSaprobicTrametes sp. 2Solitary to gregariousSaprobicPsathyrellaceaeC. disseminatusGregarious in clustersNutraceutical (Novakovic et al., 2016)P. plicalitisGregarious in clustersSaprobic (Kuo, 2015)Psathyrella spGregariousSaprobic (Kuo, 2015)	L. elegans	Solitary	Saprobic
M. xanthopusSolitary to gregariousNonedible (Tapwal et al., 2013)P. sanguineusSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitaryTherapeutic (Awala & Ayetayo,2015)T. gibbosaSolitary to gregariousMedicinal (Ren et al., 2006)T. hirsutaSolitaryMedicinal (Knezević et al., 2018)T. versicolorGregariousMedicinal (Tapwal et al., 2013)Trametes sp. 1Solitary to gregariousSaprobicTrametes sp. 2Solitary to gregariousSaprobicPsathyrellaceaeGregariousNutraceutical (Novakovic et al., 2016)P. plicalitisGregarious in clustersSaprobic (Kuo, 2015)Psathyrella spGregariousSaprobic (Kuo, 2015)	L. strigosus	Gregarious	Medicinal (Tapwal et al., 2013)
P. sanguineusSolitary to gregariousMedicinal (Cota et al., 2008)T. elegansSolitaryTherapeutic (Awala & Ayetayo,2015)T. gibbosaSolitary to gregariousMedicinal (Ren et al., 2006)T. hirsutaSolitaryMedicinal (Knezević et al., 2018)T. versicolorGregariousMedicinal (Tapwal et al., 2013)Trametes sp. 1Solitary to gregariousSaprobicTrametes sp. 2Solitary to gregariousSaprobicPsathyrellaceaeC. disseminatusGregarious in clustersNutraceutical (Novakovic et al., 2016)P. plicalitisGregarious in clustersSaprobic (Kuo, 2015)Psathyrella spGregariousSaprobic (Kuo, 2015)	M. xanthopus	Solitary to gregarious	Nonedible (Tapwal et al., 2013)
T. elegansSolitaryTherapeutic (Awala & Ayetayo,2015)T. gibbosaSolitary to gregariousMedicinal (Ren et al., 2006)T. hirsutaSolitaryMedicinal (Knezević et al., 2018)T. versicolorGregariousMedicinal (Tapwal et al., 2013)Trametes sp. 1Solitary to gregariousSaprobicTrametes sp. 2Solitary to gregariousSaprobicPsathyrellaceaeC. disseminatusGregarious in clustersNutraceutical (Novakovic et al., 2016)P. plicalitisGregarious in clustersSaprobic (Kuo, 2015)Psathyrella spGregariousSaprobic (Kuo, 2015)	P. sanguineus	Solitary to gregarious	Medicinal (Cota et al., 2008)
T. gibbosaSolitary to gregariousMedicinal (Ren et al., 2006)T. hirsutaSolitaryMedicinal (Knezević et al., 2018)T. versicolorGregariousMedicinal (Tapwal et al., 2013)Trametes sp. 1Solitary to gregariousSaprobicTrametes sp. 2Solitary to gregariousSaprobicPsathyrellaceaeC. disseminatusGregarious in clustersNutraceutical (Novakovic et al., 2016)P. plicalitisGregarious in clustersSaprobic (Kuo, 2015)Psathyrella spGregariousSaprobic (Kuo, 2015)	T. elegans	Solitary	Therapeutic (Awala & Ayetayo,2015)
T. hirsutaSolitaryMedicinal (Knezević et al., 2018)T. versicolorGregariousMedicinal (Tapwal et al., 2013)Trametes sp. 1Solitary to gregariousSaprobicTrametes sp. 2Solitary to gregariousSaprobicPsathyrellaceaeC. disseminatusGregariousNutraceutical (Novakovic et al., 2016)P. plicalitisGregarious in clustersSaprobic (Kuo, 2015)Psathyrella spGregariousSaprobic (Kuo, 2015)	T. gibbosa	Solitary to gregarious	Medicinal (Ren et al., 2006)
T. versicolorGregariousMedicinal (Tapwal et al., 2013)Trametes sp. 1Solitary to gregariousSaprobicTrametes sp. 2Solitary to gregariousSaprobicPsathyrellaceaeEndicinatusGregariousP. plicalitisGregarious in clustersSaprobic (Kuo, 2015)Psathyrella spGregariousSaprobic (Kuo, 2015)	T. hirsuta	Solitary	Medicinal (Knezević et al., 2018)
Trametes sp. 1Solitary to gregariousSaprobicTrametes sp. 2Solitary to gregariousSaprobicPsathyrellaceaeEndSaprobicC. disseminatusGregariousNutraceutical (Novakovic et al., 2016)P. plicalitisGregarious in clustersSaprobic (Kuo, 2015)Psathyrella spGregariousSaprobic (Kuo, 2015)	T. versicolor	Gregarious	Medicinal (Tapwal et al., 2013)
Trametes sp. 2Solitary to gregariousSaprobicPsathyrellaceaeC. disseminatusGregariousNutraceutical (Novakovic et al., 2016)P. plicalitisGregarious in clustersSaprobic (Kuo, 2015)Psathyrella spGregariousSaprobic (Kuo, 2015)	<i>Trametes</i> sp. 1	Solitary to gregarious	Saprobic
PsathyrellaceaeNutraceutical (Novakovic et al., 2016)C. disseminatusGregarious in clustersP. plicalitisGregarious in clustersSaprobic (Kuo, 2015)Psathyrella spGregariousSaprobic (Kuo, 2015)	<i>Trametes</i> sp. 2	Solitary to gregarious	Saprobic
C. disseminatusGregariousNutraceutical (Novakovic et al., 2016)P. plicalitisGregarious in clustersSaprobic (Kuo, 2015)Psathvrella spGregariousSaprobic (Kuo, 2015)	Psathyrellaceae		
P. plicalitisGregarious in clustersSaprobic (Kuo, 2015)Psathvrella spGregariousSaprobic (Kuo, 2015)	C. disseminatus	Gregarious	Nutraceutical (Novakovic et al., 2016)
Psathyrella sp. Gregarious Saprobic (Kuo. 2015)	P. plicalitis	Gregarious in clusters	Saprobic (Kuo, 2015)
	<i>Psathyrella</i> sp.	Gregarious	Saprobic (Kuo, 2015)
P. candolleana Solitary Saprobic	P. candolleana	Solitary	Saprobic

igodol

Russulaceae Russula sp. Schizophyllaceae	Solitary to gregarious	Saprobic
S. commune	Gregarious	Nutraceutical (Kuo, 2015)
Tricholomataceae		
<i>Mycena</i> sp	Solitary	Saprobic
P. mitis	Gregarious	Saprobic (Kuo, 2015)
M. ramealis	Gregarious in clusters	Saprobic
Xylariaceae		
D. concentrica	Gregarious	Medicinal (Dutta & Acharya, 2014)
X. papulis	Gregarious in clusters	Saprobic

Table 1 (continued)

Conclusion

In conclusion, 37 macrofungi belonging to 16 families, 26 genera, and 29 species were collected and identified. Twenty-one were saprobic, 10 medicinal, 2 nutraceutical, 1 therapeutic, and 1 for bioremediation. Generally, Paracelis in Mountain Province is a natural habitat of several mushrooms with great potential for cultivation that could demonstrate different biological activities. Therefore, further assessment of the area is recommended.

Acknowledgements

We would like to thank the people of Paracelis, Mountain Province, for letting us collect samples in the area. We are also grateful to the people who helped to make this paper came into existence: Agana Reyn Dulay, Alicia Leecel Villanueva, Emerson Diego, Lois Kristien Domingo, Larny Martin, and Jessa Mae Domingo.

References

- Adarsh, C. K., Vidyasagaran, K., & Genesh, P. N. (2018). Distribution of polypores along the altitudinal gradients in Silent Valley National Park, Southern Western Ghast, Kerala, India. *Current Research in Environmental and Applied Mycology 8*(3), 380–403.
- Baldic, A. T. (2016). Documnetation of Agroforestry Systems and Practices in Mountain Province. *Mountain Province State Polytecnic College (MPSPC) Journal 3.*
- Borchers, A. T., Keen, C. L., & Gershwin ME. (2004). Mushrooms, tumors, and immunity: an update. *Exp. Biol. Med. 229*(5), 393–406.
- Canalita, D.C., & Bajo, L.M. (2017). Toxicity, antimicrobial, and antimutagencicity potentials of the ethyl acetate extract from the mushroom *Coriolus versicolor* (L.: Fries.) Quelet. Analysis 10, 1-5.

Chang, R. (1996). Functional properties of edible mushrooms. Nutritional Revision, 54(11), 91.

- Chang, S.T., & Miles, P.G. (1992). Mushroom biology—a new discipline. *The Mycologist 6*, 64–65.
- Clausen-Heilmann, J., & Christensen, M. (2005). Wood-inhabiting macrofungi in Danish beech-forests conflicting diversity patterns and their implications in a conservation perspective. *Biological Conservation 122*, 633–642.
- Corazon, D., & Licyayo, M. (2018). Gathering practices and actual use of wild edible mushrooms among ethnic groups in the Cordilleras, Philippines. In Diversity and change in food wellbeing: Cases from Southeast Asia and Nepal. *Wageningen Academic Publishers 3*(4), 71–86.
- Cordillera Administrative Region. (2017). Cordillera Regional Development Plan 2017–2022. National Economic and Development Authority–CAR.
- De Leon, A.M., Cruz, A.S., Evangelista, A.B.B., Miguel, C.M., Pagoso, E.J.A., dela Cruz, T.E.E., Nelsen, D.J., & Stephenson, S.L. (2019). Species Listing of Macrofungi Found in the Ifugao Indigenous Community in Ifugao Province, Philippines. *Philippine Agricultural Scientist 102*(2),118–131
- De Leon, A.M., Luangsa-Ard, J.J.D., Karunarathna, S.C., Hyde, K.D., Reyes, R.G., & dela Cruz, T.E.E. (2013). Species listing, distribution, and molecular identification of macrofungi in six Aeta tribal communities in Central Luzon, Philippines. *Mycosphere* 4(3), 478– 494.
- Deocaris, C.C., de Castro, M.C.P., Oabel, A.T., Co, E., & Mojica, E.R.E. (2005). Screening for Anti-angiogenic Activity in Shiitake Mushroom (*Lentinus edodes* Berk) Extracts. *Journal* of Medical Science 5(1), 43–46.
- Department of Interior and Local Government in Cordillera Administrative Region (2015a). Municipality of Paracelis. Retrieved from the http://www.dilgcar.com/index.php/2015-07-10-09-39-31/municipality-of-paracelis.
- Department of Interior and Local Government in Cordillera Administrative Region (2015b). Province of Mountain Province. Retrieved from the http://www.dilgcar.com/index.php/2015-07-10-09-39-31/province-of-mt-province.
- Dulay, R.M.R., & Maglasang, C.C. (2017). Species Listing Of Naturally Occurring Mushrooms In Agro-Ecosystem Of Barangay Bambanaba, Cuyapo, Nueva Ecija, Philippines. International Journal of Biology, Pharmacy and Allied Sciences 6(8), 1459–1472.
- Eguchi, F., Dulay, R.M., Kalaw, S.P., Yoshimoto, H., Miyazawa, N., Seyama, T., & Reyes, R.G. (2014). Antihypertensive activities of a Philippine wild edible white rot fungus (*Lentinus sajor-caju*) in spontaneously hypertensive rats as models. *Advance Environmental Biology 8*(24), 74–81.
- Gaylan, C.M., Estebal, J.C., Tantengco, O.A., Ragragio, E.M., & Tantengco, E.M. (2018). Anti-Staphylococcal and Antioxidant Properties of Crude Ethanolic Extracts of Macrofungi Collected from the Philippines. *Pharmacological Jornal 10*(1), 106–109.

- Gibertoni, T.B., Medeiro, P.S., Soare, A.M.S., Gomes-Silva, A.C., Santos, P.J.P., Satăo, H.M.P., & Ferreira, L.V. (2016). The distribution of polypore fungi in endemism centres in Brazilian Amazonia. *Fungal Ecology*, *20*, 1–6.
- Gilbert, GS. (2005). The dimension of plant disease in the tropical forests. *In* Biotic interactions in the tropics. Edited by Burslem, D.R.F.P., Pinard, M. A., & Hartley, S. Cambridge University Press 141–164.
- Gilbert, G.S., & Sousa, W.P. (2002). Host specialization among wood-decaying polypore fungi in a Caribbean mangrove forest. Biotropica 34, 396–404.
- Hawksworth, DL. (2001). The magnitude of fungal diversity: The 1.5 million species estimate revisited. *Mycological Research*, 105, 1422–1432.
- Kalač, P. (2013). A review of chemical composition and nutritional value of wild-growing and cultivated mushrooms. Journal of Science and Food Agriculture 93(2), 209–218.
- Kuo, M. (2015). Mushroom taxonomy: The big picture. Retrieved from the MushroomExpert.com. http://www.mushroomexpert.com/taxonomy.html.
- Laessoe, L., & Lincoff, G. (1998). Mushrooms (Eyewitness Handbooks) Kyodo printing Co. Singapore. 303 P.
- Liwanag, J.M.G., Santos, E.E., Flores, F.R., Clemente, R.F., & Dulay, R.M.R. (2017). Species Listing of Macrofungi in Angat Watershed Reservation, Bulacan Province, Luzon Island, Philippines. International Journal of Biology, Pharmacy and Allied Science 6 (5), 1060–1068.
- Lodge, D.J., Ammiranti, O'dell, T.E., & Mueller, G.M. (2004). Collecting and discribing macrofungi in: biodiversity of fungi: inventory and monitoring methods (eds Mueller GM, Bills GF, Foster MS). *Elsevier Academic Press*, USA. 128–158.
- López-Quintero, C.A., Straatsma, G., Franco-Molano, A.E., & Boekhout, T. (2012). Macrofungal diversity in Colombian Amazon forests varies with regions and regimes of disturbance. *Biodiversity Conservation 21*(9), 2221–2243.
- May, R. (1991). A fondness for fungi. Nature (London), 352, 475-476.
- Mueller, G.M., Schmit, J.P., Leacock, P.R., Buyck, B., Cifuentes, J., Desjardin, D.E., & Lodge, D.J.
 (2007) Global diversity and distribution of macrofungi. *Biodiversity and conservation* 16(1), 37–48.
- Niem, J.M., & Baldovino, M.M. (2015). Initial Checklist of Macrofungi in the Karst Area of Cavinti, Laguna. Museum Publications in Natural History 4(1), 11–25.
- Orgiazzi, A., Bardgett, R.D., Barrios, E., Behan-Pelletier, V., Briones, M.J.I., Chotte, J.L., De Deyn, G.B., Eggleton, P., Fierer, N., Fraser, T., Hedlund, K., Jeffery, S., Johnson, N.C., Jones, A., Kandeler, E., Kaneko, N., Lavelle, P., Lemanceau, P., Miko, L., Montanarella, L., Moreira, F.M.S., Ramirez, K.S., Scheu, S., Signh, B.K., Six, J., van der Putten, W.H., & Wall, D.H. (2016). Global soil biodiversity Atlas. *European Union*, 176.

Vol. 5 No. 2 (August 2021) ISSN: 2507-9638 DOI: http://dx.doi.org/10.22137/v5n2.03

- Parveen, A., Khataniar, L., Goswami, G., Hazarika, D.J., Das, P., Gautom, T., Barooah, M., & Boro, R.C. (2017). A Study on the Diversity and Habitat Specificity of Macrofungi of Assam, India. International Journal of Current Microbiology and Applied Sciences 6(12), 275–297.
- Phillips, R. (2006). Mushroom of North America. Little, Brown and Company (Canada) Limited, 310.
- Quimio, TH. (2001). Common mushroom in Mt. Makiling. University of the Philippines, Los Baňos, Laguna. *Museum of National History*, 8, 83.
- Reyes, R.G., Del Rosario, M.A.G., Padua, J.P.G., Malonzo, M.A.C., Barza, A.J.J., Sumi, R., Mori, N., Miyazawa, N., & Eguchi, F. (2017) The first report on the mycelial growth performance and antibacterial activity of *Collybia reinakeana* RGR-FE–NSC strain, a Philippine endemic edible mushroom. *International Journal of Pharmacology* 6(1), 113–119.
- Rossman, A. (1994). A strategy for an all taxa inventory of fungal biodiversity. In: Peng CI, Chou Ch (eds), Biodiversity and terrestrial ecosystem. *Academia Sinica Monograph*, 14, 169–194.
- Rydin, H., Diekmann, M., & Hallingback, T. (1997). Biological Chracteristics, Habitat
 Associations, and Distribution of Macrofungi in Sweden. *Conservation Biology*, 11(3), 628–640.
- Sibounnavong, P., Divina, C.C., Kalaw, S.P., Reyes, R.G., & Soytong, K. (2008). Some species of macrofungi at Puncan, Carranglan, Nueva Ecija in the Philippines. *Journal of Agricultural Technology* 4, 105–115.
- Schmitt, J.P. (2005). Species richness of tropical wood-inhabiting macrofungi provides support for species-energy theory. *Mycologia* 97, 751–761
- Tadiosa, E.R., Agbayani, E.S., & Agustin, N.T. (2011) Preliminary Study on the Macrofungi of Bazal-Baubo Watershed, Aurora Province, Central Luzon, Philippines. Asian Journal of Biodiversity 2, 149–171.
- Tadiosa, E.R., Arsenio, J.J., & Marasigan, M.C. (2007). Macroscopic fungal diversity of Mt. Makulot, Cuenca, Batangas, Philippines. *Journal of Nature Studies* 6, 1–2.
- Zhang, Y., Hyde, K.D., Zhou, D.Q., Zhao, Q., & Zhuo, T.X. (2010). Diversity and ecological distribution of macrofungi in the Laojun Mountain region, southwestern China. *Biodiversity and Conservation* 16, 3545–3563.